

Primary Amebic Meningoencephalitis Caused by *Naegleria fowleri*, Karachi, Pakistan

Sadia Shakoor, Mohammad Asim Beg, Syed Faisal Mahmood, Rebecca Bandea, Rama Sriram, Fatima Noman, Farheen Ali, Govinda S. Visvesvara, and Afia Zafar

We report 13 cases of *Naegleria fowleri* primary amebic meningoencephalitis in persons in Karachi, Pakistan, who had no history of aquatic activities. Infection likely occurred through ablution with tap water. An increase in primary amebic meningoencephalitis cases may be attributed to rising temperatures, reduced levels of chlorine in potable water, or deteriorating water distribution systems.

Primary amebic meningoencephalitis (PAM) is a fatal disease caused by the thermotolerant free-living ameba *Naegleria fowleri*. Found worldwide in moist soil and freshwater, these amoebae proliferate during summer when ambient temperature increases. The organism enters the nasal cavity when water contaminated with amoebae is aspirated. Subsequently, it invades the central nervous system through the olfactory neuroepithelium and causes a fatal infection that clinically resembles acute bacterial meningitis. We report 13 cases of *N. fowleri* PAM in a period of 17 months in the coastal city of Karachi, Pakistan.

The Study

In June 2008, a 30-year-old, previously healthy man was referred to the Aga Khan University Hospital with a 2-day history of high-grade fever, severe headache, and seizures. He was comatose with a fixed and dilated left eye pupil. Magnetic resonance imaging showed basal meningeal enhancement. A lumbar puncture found an opening pressure of 44 cm H₂O. Cerebrospinal fluid (CSF) analysis showed low glucose (<5 mg/dL), high protein level (1,028 mg/dL), and lymphocytic pleocytosis (900 cells/mm³ with 85% lymphocytes). Gram stain and latex agglutination

(LA) test results were negative for bacteria. Wet film of CSF showed motile amebic trophozoites, but CSF volume was insufficient for amebic culture. A preliminary diagnosis of PAM was made, and therapy was begun with intravenous amphotericin B plus oral rifampin and fluconazole. The patient died 4 days after admission.

In September 2008, a previously healthy 25-year-old man was admitted with a 24-hour history of fever, vomiting, and neck rigidity. CSF analysis indicated hypoglycorrhachia, elevated protein level, and neutrophilic pleocytosis. Gram stain and LA test results were negative for bacteria, and wet preparation of CSF showed motile amebic trophozoites. Ptosis of the left eye and a fixed dilated pupil developed on the day of admission, and he was intubated for airway protection. Despite therapy with intravenous amphotericin B, oral fluconazole, and rifampin, his condition deteriorated, and he died 14 days after admission. CSF was cultured on nonnutrient agar on a lawn of *Escherichia coli* American Type Culture Collection (Manassas, VA, USA) 29522 in Page ameba saline (1). Cultured amoebae produced flagellates in distilled water at 37°C within 15–30 min.

On the basis of this apparent upsurge of cases, a laboratory policy was instituted at the Aga Khan University Hospital to perform wet mounts of all processed CSF samples that were consistent with bacterial meningitis but had negative Gram stain and LA test results. Although no new cases of PAM were detected in 2008, 11 were identified from April through November 2009. Case-patients were referred from tertiary-care hospitals in Karachi (Table).

Twelve of 13 patients were male; ages ranged from 16 to 64 years (mean \pm SD 31.0 \pm 15.33 years). All were residents of Karachi but lived in different districts (online Appendix Figure, www.cdc.gov/EID/content/17/2/256-appF.htm). Only 1 patient acknowledged a history of swimming. All patients' conditions were treated with amphotericin B (1.5 mg/kg/d) and rifampin (600 mg/d). Most patients also received either fluconazole or itraconazole. All required intubation and ventilation within 24 hours of hospital admission, and treatment for PAM was started within 48–72 hours of admission. Nonetheless, only 2 patients survived for >5 days after admission; the mean \pm SD time from symptom onset to death was 6.38 \pm 3.15 days (range 3–15 days).

CSF samples from all patients (except the first) were positive for amebic culture. Furthermore, CSF samples collected from 3 patients in July 2009 were confirmed by real-time PCR for *N. fowleri* DNA at the Division of Parasitic Diseases, Centers for Disease Control and Prevention, Atlanta, Georgia, USA. Briefly, primers NaegIF192 (3'-GTG CTG AAA CCT AGC TAT TGT AAC TCA GT-5') and NaegIR344 (5'-CAC TAG AAA AAG CAA ACC TGA AAG G-3') were used to amplify a 153-bp fragment, detected by the hexachlorofluorescein (HEX)-labeled probe NfowIP

Author affiliations: Aga Khan University Hospital, Karachi, Pakistan (S. Shakoor, M.A. Beg, S.F. Mahmood, F. Ali, A. Zafar); Centers for Disease Control and Prevention, Atlanta, Georgia, USA (R. Bandea, R. Sriram, G.S. Visvesvara); and Liaquat National Hospital, Karachi (F. Noman)

DOI: 10.3201/eid1702.100442

Table. Clinical features and CSF analysis results for 13 patients with *Naegleria fowleri* meningoencephalitis, Karachi, Pakistan, 2008–2009*

Characteristic	Patient nos., in order of treatment													Descriptive statistics†
	1	2	3	4	5	6	7	8	9	10	11	12	13	
Age, y	31	25	30	36	60	30	64	18	22	16	18	18	35	Mean 31.0 ± 15.33
Date treatment sought	2008 Jul	2008 Sep	2009 Apr	2009 May	2009 Jul	2009 Jul	2009 Jul	2009 Jul	2009 Aug	2009 Aug	2009 Oct	2009 Oct	2009 Nov	
Duration of illness before seeking treatment, d	2	1	2	4	4	1	2	5	2	2	2	3	3	Mean ± SD 2.5 ± 1.19
Duration of survival after symptom onset, d	6	15	3	6	7	9	7	6	5	3	3	7	6	Mean ± SD 6.38 ± 3.15
Fever	+	+	+	+	+	+	+	+	+	+	+	+	+	Present in 13/13 (100%)
Headache	+	+	+	+	+	+	+	–	+	+	+	+	+	Present in 12/13 (92.3%)
Seizures	+	–	+	–	+	–	+	–	+	+	+	+	–	Present in 8/13 (61.5%)
CSF analysis														
Glucose, mg/dL‡	<5	42	2	<5†	26	<5†	<5‡	30	<5‡	<5‡	80	50	<5‡	Undetectable in 53.8%
Protein, mg/dL	1,028	418	909	774	504	1,147	1342	320	998	330	371	179	1,296	Mean ± SD 739.69 ± 405.29
Leukocytes, cells/mm ³	900	900	5,976	2,000	2,500	7,500	5,200	6,000	840	6,500	150	185	11,750	Mean ± SD 3,877.0 ± 3,565.37
Neutrophils, %	15	90	90	90	60	80	90	90	90	96	75	25	95	Neutrophilic pleocytosis, 84.6
Lymphocytes, %	85	10	10	10	40	20	10	10	10	4	25	75	5	Lymphocytic pleocytosis, 15.4
CSF amebic culture	NA	+	+	+	+	+	+	+	+	+	+	+	+	NA
CSF PCR for <i>N. fowleri</i>	NA	NA	NA	NA	+	+	+	NA	NA	NA	NA	NA	NA	NA

*All patients were male except patient no. 11, who was female. CSF, cerebrospinal fluid; +, present; NA, no data available.

†Statistics derived in SPSS software version 16.0 (LEAD Technologies, Charlotte, NC, USA).

‡Glucose <5 mg/dL = undetectable.

(5'-HEX-AT AGC AAT ATA TTC AGG GGA GCT GGG C-BHQ1-3'). PCR was performed in an Mx3000P real-time thermocycler (Stratagene, La Jolla, CA, USA), with 2 initial incubations at 50°C for 2 min (incubation for uracil-DNA-glycosylase activity) and 95°C for 2 min (activation of Platinum *Taq* DNA-polymerase), respectively, followed by 40 cycles of 95°C for 15 s and 63°C for 60 s. Fluorescence was measured after each 63°C incubation. Results were analyzed by using Mx3000P version 2.0 software (2).

Domestic tap water was obtained for amebic culture from 2 patients' homes (second and seventh patients in the series). Water samples (100 mL) were passed through a Millipore filter (Millipore Corp., Billerica, MA, USA), which was then inoculated face down on a nonnutrient agar plate with a lawn of *E. coli* as described above. Amebae

were isolated from both water samples. However, only cultured amebae from the seventh patient's sample were analyzed by real-time PCR, and *N. fowleri* DNA was detected in the sample.

Conclusions

We report an increase of *N. fowleri* PAM cases in Karachi, Pakistan. Most cases were in healthy young adults with acute, fatal meningitis. Although enhanced case detection after instituting measures to improve the diagnosis of PAM may have contributed toward the rise in cases, sporadic previous reports from this region indicate that this is not the sole reason (3,4).

PAM is associated with freshwater swimming (5), and outbreaks have also been associated with poorly chlorinat-

ed swimming pools (6). However, all but 1 of the patients in this case series denied recent freshwater swimming or recreational water activities. Nevertheless, because all patients were Muslim, they routinely performed ritual ablution, which involves taking water into the nostrils. Infection acquired through this route has been reported (7). Because patients used domestic tap water for ablutions, we tested water from 2 patient's homes and found *N. fowleri* amoebae by culture. Additionally, *N. fowleri* DNA was identified by real-time PCR in a water sample from 1 patient's home. PAM, resulting from aspiration of untreated ground water containing *N. fowleri* amoebae, has been reported in 2 children from Arizona (8).

The presence of *N. fowleri* DNA in the ground water supply has been described (9). *N. fowleri* infections have also been described in children from Australia after exposure to water transported through overland pipes (10). The presence of *N. fowleri* amoebae in Karachi's municipal water supply may have several explanations. Karachi lacks an indigenous water source, and water obtained from 2 suburban freshwater lakes is not adequately filtered or chlorinated (11). Moreover, frequent leaks in water and sewage pipes can cause seepage of sewage into the water supply (12), which may be a potential reservoir for *N. fowleri* amoebae (13).

Demonstrating *N. fowleri* amoebae in the water supply, however, does not explain the sudden increase in the number of cases. *N. fowleri* amoebae are thermotolerant, and outbreaks have been linked to steep rises in temperature (14). Recent temperature records from Karachi have also shown a temperature surge, and a further rise is expected in future summers (15). Our patients were also exposed to higher average monthly temperatures (Figure). Several factors may have precipitated this outbreak: an increase in the number of pathogens (due to rising temperatures), changes in the ecosystem of the lakes, and further deterioration in the quality

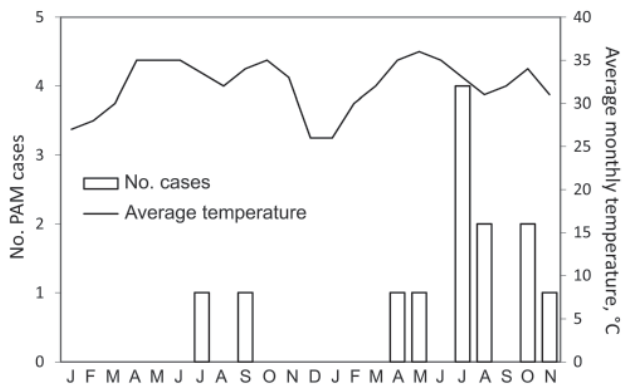


Figure. Primary amebic meningoencephalitis (PAM) cases seen in 2008 and 2009 by month (white bars) and average monthly peak temperatures (black line), Karachi, Pakistan. Karachi monthly temperatures obtained from <http://www.wunderground.com/global/PK.html>.

of water treatment and distribution systems.

This report highlights the emergence of fatal *N. fowleri* infection in a megacity. Changing climatic conditions may have contributed toward this sudden upsurge, which has serious consequences for the public at large. Urgent epidemiologic investigation into relevant environmental factors is needed to identify reasons for this sudden rise in PAM cases.

Acknowledgments

We thank Maqboola Dojki and Noureen Saeed for assistance with managerial responsibilities.

Financial support for shipping materials to the Centers for Disease Control and Prevention was provided by the Deanery, Aga Khan University Hospital, Karachi.

Dr Shakoor is a clinical microbiologist in training at the Aga Khan University Hospital, Karachi, Pakistan. Her research interests include tuberculosis and emerging parasitic diseases.

References

1. Isenberg HD, ed. Clinical microbiology procedures handbook. 2nd ed. Washington: American Society for Microbiology; 2004.
2. Qvarnstrom Y, Visvesvara GS, Sriram R, Da Silva AJ. Multiplex real-time PCR assay for simultaneous detection of *Acanthamoeba* spp., *Balamuthia mandrillaris*, and *Naegleria fowleri*. *J Clin Microbiol*. 2006;44:3589–95. DOI: 10.1128/JCM.00875-06
3. Jamil B, Ilyas A, Zaman V. Primary amoebic meningoencephalitis. *Infectious Disease Journal of Pakistan*. 2008;17:66–8 [cited 2010 Dec 20]. <http://www.idspak.org/journal/2008/april-june/page66-68.pdf>
4. Ahmed K, Dil AS, Ahmed I, Hussain S, Lakhnana NK. Primary amoebic meningoencephalitis: report of a case. *Int J Pathol*. 2007;5 [cited 2010 Dec 20]. [http://www.jpathology.com/Issues/Previous%20Articles/8\(9\).html](http://www.jpathology.com/Issues/Previous%20Articles/8(9).html)
5. Craun GF, Calderon RL, Craun MF. Outbreaks associated with recreational water in the United States. *Int J Environ Health Res*. 2005;15:243–62. DOI: 10.1080/09603120500155716
6. Shenoy S, Wilson G, Prashanth HV, Vidyakshmi K, Dhanashree B, Bharath R. Primary Meningoencephalitis by *Naegleria fowleri*: first reported case from Mangalore, South India. *J Clin Microbiol*. 2002;40:309–10. DOI: 10.1128/JCM.40.1.309-310.2002
7. Lawande RV, Macfarlane JT, Weir WR, Awunor-Renner C. A case of primary amoebic meningoencephalitis in a Nigerian farmer. *Am J Trop Med Hyg*. 1980;29:21–5.
8. Okuda DT, Hanna HJ, Coons SW, Bodensteiner JB. *Naegleria fowleri* hemorrhagic meningoencephalitis: report of two fatalities in children. *J Child Neurol*. 2004;19:231–3.
9. Marciano-Cabral F, MacLean R, Mensah A, LaPat-Polasko L. Identification of *Naegleria fowleri* in domestic water sources by nested PCR. *Appl Environ Microbiol*. 2003;69:5864–9. DOI: 10.1128/AEM.69.10.5864-5869.2003
10. Dorsch MM, Cameron AS, Robinson BS. The epidemiology and control of primary amoebic meningoencephalitis with particular reference to South Australia. *Trans R Soc Trop Med Hyg*. 1983;77:372–7. DOI: 10.1016/0035-9203(83)90167-0
11. Dawn NEWS Internet Edition. 'Contaminated water' being supplied to city. 2009 July 22 [cited 2010 Dec 20]. http://www.dawn.com/fixd/arch/arch_2009.html

12. Dawn NEWS Internet Edition. Contaminated water blamed for gastro in Landhi. 2009 July 17 [cited 2010 Dec 20]. http://www.dawn.com/fixd/arch/arch_2009.html
13. Lawande RV, Ogunkanmi AE, Egler LJ. Prevalence of pathogenic free-living amoebae in Zaire, Nigeria. *Ann Trop Med Parasitol*. 1979;73:51–6.
14. Cogo P E, Scaglia M, Gati S, Rosetti F, Alaggio R, Laverda AM, et al. Fatal *Naegleria fowleri* meningoencephalitis, Italy. *Emerg Infect Dis*. 2004;10:1835–7.
15. Sajjad SH, Hussain B, Khan MA, Raza A, Zaman B, Ahmed I. On rising temperature trends of Karachi in Pakistan. *Clim Change*. 2009;96:539–47. DOI: 10.1007/s10584-009-9598-y

Address for correspondence: Syed Faisal Mahmood, Department of Medicine, Aga Khan University Hospital, Karachi 75800, Pakistan; email: faisal.mahmood@aku.edu

etymologia

Naegleria fowleri

[nə'gliəriə fau(ə)l'ər-i]

From F.P.O. Nägler, an early 20th century bacteriologist, and Malcolm Fowler, an Australian physician. In 1912, A. Alexeieff proposed a new genus for questionable amoeboid forms, which he named Nägleria in honor of Nägler's work in identifying amoebae that pass through a biflagellate stage. Fifty-three years later, a report from Australia described human meningoencephalitis caused by an amebo-flagellate, later recognized as a member of *Naegleria*. In 1970, the pathogen was designated *Naegleria fowleri* after Fowler, who obtained one of the first isolates from human brain tissue.

Source: Alexeieff A. Sur les caractères cytologiques et la systématique des amibes du groupe limax (*Naegleria* nov gen et *Hartmannia* nov. gen) et des amibes parasites des vertèbres (*Proctamoeba* nov. gen). *Bull de la Soc Zool de France*. 1912; 37:55; Calkins GN. Genera and species of amoeba. In: *Transactions of the Fifteenth International Congress on Hygiene and Demography*, Vol. II, Washington, September 23–28, 1912. Washington: General Printing Office; 1913; Fowler M, Carter RF. Acute pyogenic meningitis probably due to *Acanthamoeba* sp.: a preliminary report. *BMJ*. 1965;2:740–2. DOI: 10.1136/bmj.2.5464.734-a; Marciano-Cabral F. Biology of *Naegleria* spp. *Microbiol Rev*. 1998;52:114–33.

SUBSCRIBE

EMERGING INFECTIOUS DISEASES®

YES, I would like to receive Emerging Infectious Diseases. Please add me to your mailing list.

Return:

Email:
eideditor@cdc.gov

Fax: 404 639-1954

Mail to:

EID Editor
CDC/NCID/MS D61
1600 Clifton Rd, NE
Atlanta, GA 30333

Number on mailing label:(required) _____

Name: _____

Full mailing address: (BLOCK LETTERS)

Full text free online at www.cdc.gov/eid